## **COMMENTARY**

## NOVEL THERAPEUTICS ACTING VIA PURINE RECEPTORS

KENNETH A. JACOBSON,\*† BHARAT K. TRIVEDI,‡ PAUL C. CHURCHILL§ and MICHAEL WILLIAMS||

\*Laboratory of Bioorganic Chemistry, NIDDK, National Institutes of Health, Bethesda, MD 20892; ‡Warner-Lambert Co., Pharmaceutical Research Division, Ann Arbor, MI 48105; \$Department of Physiology, Wayne State University, Detroit, MI 48201; and |Neuroscience Research, Abbott Laboratories, Abbott Park, IL 60064, U.S.A.

A recent conference entitled Purines in Cell Signalling: Targets for New Drugs, held in Rockville, Maryland, in September, 1989, was one indication of the increasing interest in developing agonists and antagonists of  $P_1$ -(adenosine) and  $P_2$ -(ATP) purinoceptors [1] as potential therapeutic agents. Extracellular adenosine, acting at its membrane bound  $A_1$  and  $A_2$  receptors, is a ubiquitous modulator of cellular activity. The purine can arise from several sources including ATP hydrolysis by ectokinase activity in the region of the nerve terminal [2] and from S-adenosylhomocysteine [3] and ATP within the cell. Together with its more stable analogs, adenosine is a potent inhibitor of neurotransmitter release in both the central and peripheral nervous systems, and in cardiac, adipose and other tissues. Adenosine can also affect blood pressure and heart rate as well as modulate the function of the immune, inflammatory, gastrointestinal, renal and pulmonary systems, either via its effects on transmitter release or directly via receptor mechanisms altering intracellular transduction processes.

#### Adenosine agonists

The first reported physiological action of adenosine (1) was in regard to its ability to lower blood pressure, a finding reported by Drury and Szent-Györgyi in 1929 [4]. This seminal article stimulated interest in the potential use of adenosine as an antihypertensive agent. However, clinical evaluation of the purine in the 1930s as a hypotensive agent was ephemeral due to its poor efficacy, a result of a short half-life, and led to a degree of negativity about its therapeutic potential [5]. Interestingly, the recently approved use of adenosine as an agent for the treatment of supraventricular tachycardia [6], developed by Berne and coworkers, is successful primarily because of the short half-life of the native compound.

In the 60 years since the work of Drury and Szent-Györgyi, many other uses for adenosine agonists have been proposed (Table 1). These actions are dependent almost exclusively on the degree of selectivity of the ligands for either  $A_1$  or  $A_2$  receptor

subtypes and the present knowledge related to the physiological role(s) of these receptors.

Several thousand potent adenosine agonists (selected structures in Table 2) have been synthesized in industrial and other medicinal chemistry laboratories in the past 40 years [1]. Anecdotal reports of limited human trials in the early 1950s with 2chloroadenosine (2-CADO, 3) suggested that the longer half-life of this agonist and its greater potency led to an unacceptable, near fatal drop in blood pressure that precluded further testing. In the 1970s, Boehringer-Ingelheim conducted a major program synthesizing  $N^6$ -alkyl substituted adenosine analogs [including  $N^6$ -(R)-phenyliso-propyladenosine, R-PIA, 6] as potential hypotensive agents [17]. The vasodilatory potency of such agonists was improved by Abbott Laboratories with the synthesis of 5'-N-ethylcarboxamidoadenosine [18] (NECA, 2), which is still one of the more potent agonists at the A<sub>2</sub> adenosine receptor subtype. Interestingly, the original patent for this compound [18] described it as a rodent poison. Takeda Pharmaceuticals discovered that arylamino substitution at the C-2 position of adenosine resulted in potent vasodilators among which, CV 1808 [2-(phenylamino)adenosine, 4] was later found to be a weak  $(K_i = 100 \text{ nM})$  but 7-fold A<sub>2</sub>-selective adenosine agonist [20].

One of the most significant efforts in the medicinal chemistry of adenosine derivatives was that of Parke-Davis in the 1980s [10, 20-23]. This program encompassed both agonist and antagonist pharmacophores for adenosine receptors and resulted in the identification of several classes of  $N^6$ -substituted adenosine derivatives. The majority of these were  $A_1$  selective, but others including CI-936 [ $N^6$ -(2,2diphenylethyl)adenosine, 7] [10] were potent agonists at A2-adenosine receptors. Continued efforts to optimize the A<sub>2</sub> affinity of this series of compounds resulted in a series of substituted diphenylethyladenosine derivatives, from which DPMA  $(N^{\delta}$ - [2-(3,5-dimethoxyphenyl)-2-(2-methylphenyl)ethyl]adenosine, PD 125,944, 8) was identified as having a 32-fold selectivity for the A<sub>2</sub> receptor, with a  $K_i$  value of 4.4 nM [21].

Interestingly, when an anilino function was incorporated into the C2-position of DPMA, such as in the case of CV-1808, with an intent to improve the  $A_2$  potency and/or selectivity, it rendered an

<sup>†</sup> Correspondence should be addressed to: Dr. K. A. Jacobson, Bldg. 8A, Rm. B1A-17, National Institutes of Health, Bethesda, MD 20892.

Table 1. Potential therapeutic uses of drugs acting at adenosine receptors [1, 7–16]
--

Tissue/organ	Adenosine agonist	Adenosine antagonist
CNS	Antipsychotic [9, 10], anxiolytic, sedative, analgesic, neuroprotective [12], anti-convulsant, anti-hypertensive (central mech.), anti-Parkinson	Nootropic, cognition enhancer [11]
Heart	Antiarrhythmic [13], congestive heart failure	Cardiotonic
Kidneys	Antihypertensive (by inhibition of renin release [14])	Diuretic, kidney protective [14]
Vasculature	Vasodilator	_
Immune	<del>_</del>	Immunostimulant
Inflammatory	Anti-inflammatory	_
Reproductive	_	Contraceptive (lowers sperm motility)*
Gastrointestinal	Inhibitory to gastric acid secretion [15]	
Lungs	_	Anti-asthmatic [16, †]

<sup>\*</sup> Hoskins D and Vijayaraghavan S, Intracellular adenosine regulates mammalian sperm motility. Presented at *Purine Nucleotides and Nucleotides in Cell Signalling: Targets for New Drugs*, Bethesda, MD, Sept. 17, 1989, Abstr. D1.

analog (10) with significantly lower affinity at both the receptors  $(A_1 K_i = 10,300 \text{ nM}; A_2 K_i = 340 \text{ nM})$ [22]. The decrease in the binding affinity for this analog compared to the parent compound was attributed to the steric factors involved at the C2 domain of the binding site. However, modification of the 5'-hydroxymethyl function to a carboxamidoethyl function, such as in the case of NECA, provided an analog (9) with binding affinity ( $A_1 K_i = 207 \text{ nM}$ ;  $A_2$  $K_i = 5.6 \text{ nM}$ ) similar to the parent compound [21]. These data suggest that there may exist two separate binding domains at the A<sub>2</sub> receptor where these adenosine analogs could interact independently when substituted either in the  $N^6$ -position or in the 5'-position. However, for the  $A_1$  receptor, the binding affinity seems to be dependent on the interactions at both the  $N^6$  and the 5'-domain. For example, conversion of the  $A_1$ -selective agonist  $N^6$ -(2-S-endo-norbonyl)adenosine (S-ENBA, 12) into a 5'-chloro derivative (13) resulted in a compound which is one of the most potent ( $K_i = 0.24 \text{ nM}$ ) and highly selective (16,000-fold) ligands for the adenosine A<sub>1</sub> receptor [23]. Thus, selectivity could be enhanced for the A<sub>1</sub> receptor by simply modifying the 5'-position of the molecule. Indeed, in recent years, extensive work on structure-activity relationships has been carried out in various laboratories, which not only enhanced our understanding of the binding domain of these receptors but also has provided major insights into the key structural features required for better affinity and/ or selectivity at these receptors.

Hybrid modifications of the purine nucleoside pharmacophore in the 5'- and C2 positions by the CIBA-Geigy group led to over 200 highly A<sub>2</sub>-selective adenosine agonists, among which CGS

21680 [2-(2-p-carboxyethyl)phenethylamino-5'-N-ethylcarboxamidoadenosine, 5] was 140-fold selective for the  $A_2$ -adenosine receptor subtypes with an affinity of 21 nM [24]. Similar  $5'/N^6$  hybrid molecules are generally  $A_1$  selective [25].

Despite the intensive efforts on the part of many companies over the past 30 years with the attendant discovery of successive generations of potent and selective adenosine agonists, it is ironic that only adenosine itself has been approved as a therapeutic agent. Lyphomed/Medco introduced adenosine as Adenocard<sup>TM</sup> for the treatment of supraventricular tachycardia (SVT), a transitory blockade of AV node conduction leading to an attenuation of abnormal heart rhythms [6], based on work by Belardinelli, Berne, and colleagues. The crucial feature in the administration of adenosine, rather than a more stable (and potent) analog, for SVT was the very short half-life (10 sec) of the nucleoside [26]. As a result of metabolic inactivation, primarily through adenosine deaminase, the effects of intravenously-administered adenosine are limited to actions in the vascular system and, as a result, potential side-effects are avoided. Interestingly, Adenocard<sup>TM</sup> has been upgraded recently to the FDA's coveted 1A designation for a compound fulfilling unmet medical need. Medco is also pursuing use patents for diagnostic applications of adenosine in various cardiovascular disease states, while the purine is being used for controlled hypotension in aneurysm surgery in Sweden [27] and evaluated for similar use in North America.

In contrast, the majority of potent and selective adenosine analogs have remained in the research tool category. One reason for this is the perceived potential actions of nucleosides and nucleotides as

<sup>†</sup> Anti-asthmatic effects of theophylline are complex and appear to be related to inhibition of phosphodiesterases and other mechanisms.

Table 2. Structures of selected adenosine agonists and affinities at  $A_1$ - and  $A_2$ -adenosine receptors (data from references given in text)

Compound	R	R'	R"	$K_i$ A <sub>1</sub>	$K_iA_2$
1	Н	Н	CH₂OH	12.8	37
2	Н	Н	CONHEt	6.3	12
3 .	Н	Cl	CH <sub>2</sub> OH	9.3	63
4	н	ин-	CH <sub>2</sub> OH	600	116
5	н ин(сн	(CH <sub>2</sub> ) <sub>2</sub> COOH	CONHEt	2600	15
6	HXCH3	H	CH₂OH	1.17	124
7		Н	CH₂OH	6.8	25
8	осн,	н	СН₂ОН	142	4.4
9	H₃Ċ `	Н	CONHEt	207	5.6
10	**	ин-⟨т⟩	CH₂OH	10,300	340
11		Н	СН₂ОН	22	412
12	H	н	CH₂OH	0.3	1390
13	••	Н	CH₂Cl	0.24	3900

antimetabolites, which would be undesirable here. Many such compounds including the anti-AIDS drugs AZT (3-azidothymidine), ddI (2',3'-dideoxyinosine) and ddC (2',3'-dideoxycytosine) are effective antiviral agents and have the potential both to interfere

with nucleic acid replication and to suppress the immune response. Another issue relates to the ubiquitous actions of the purine nucleoside. Because adenosine receptors, both  $A_1$  and  $A_2$ , are present in nearly all tissues, it is probable that agents acting

via activation or inhibition of such receptors will have a multitude of actions. Xanthine adenosine antagonists (see below) with potential as central stimulants may also function as cardiotonic agents [28]. Conversely, agonists affecting cardiovascular function will cause sedation in the CNS and also alter renal function. This liability for side-effects, perceived or real, is a major barrier to the use of synthetic adenosine agonists, which are potent and relatively metabolically stable *in vivo*.

For example, Parke-Davis sought to develop for human therapeutic use several promising adenosine agonists, including CI-936 (7) evaluated as an antipsychotic [9]. CI-936 exhibited antipsychotic-like activity in mice by inhibiting motor activity (ED<sub>50</sub> = 1.3 mg/kg) without producing significant ataxia  $(ED_{50} = 145 \text{ mg/kg})[10]$ . Furthermore, in the Sidman avoidance paradigm, a test predictive of antipsychotic activity, this compound selectively blocked the response with an ED<sub>50</sub> of 2.5 mg/kg. Extensive toxicological evaluation in mice, rats, dogs and monkeys\* showed that CI-936 was well tolerated at daily doses up to 40 mg/kg in rats, 12.5 mg/kg in dogs, and 12.5 mg/kg in monkeys for up to 2 weeks. Sporadic emesis was noted at 12.5 mg/kg in dogs and at 25 mg/kg in monkeys.\* Unfortunately, the lack of selectivity of CI-936 for its CNS versus cardiovascular actions and other side-effects precluded further development.

Another series of novel adenosine agonists identified at Parke-Davis, the N<sup>6</sup>-(benzocycloalkyl)adenosines, had excellent blood pressure low ering activity in a spontaneously hypertensive rat model. $\dagger$  From this series of compounds, (R)- $N^6$ -(2,3-dihydro-1*H*-inden-1-yl)adenosine (11) was identified as a potential antihypertensive agent. The oral doses required to lower mean arterial blood pressure by 20% (ED<sub>20</sub>) in the spontaneously hypertensive rats (SHR), renal hypertensive rats, DOCA (deoxycorticosterone) salt hypertensive rats and renal hypertensive dogs (RHD) were 1.2, 0.3, 1.3 and 1.5 mg/kg respectively [29]. At the ED<sub>20</sub>, heart rate was elevated moderately in RHD (+30 beats/min) and decreased in SHR (-40 beats/ min). A long duration of action was observed in all hypertensive models, with effects lasting beyond 10 hr [29]. However, this compound showed significant toxicity in dogs [30] including coronary arterial lesions, endothelial swelling, disruption of the internal elastic lamina, and necrosis. It is not clear whether the toxicity observed was the consequence of the pharmacological effects exerted by the compound or was due to the inherent toxicity associated with this class of chemicals.

Even agents which are selective for the periphery or more selective for a receptor subtype would suffer from the potential multiplicity of side-effects. For most of the envisioned applications of adenosine agonists (Table 1), such effects could conceivably be as serious as the conditions for which they were intended to treat. This would be unacceptable where therapeutically effective and safe drug entities are already meeting medical needs.

Such pessimism may be balanced to some extent by the fact that chronic ingestion of the adenosine antagonist, caffeine, is rarely associated with deleterious effects except in individuals who are extremely prone to tremors and convulsions. The underlying cause of the latter response is not known nor indeed is there much information available in regard to adenosine pathophysiologies in either cardiovascular or CNS disease states for which the nucleoside has been loosely targeted. An anomaly of the drug discovery process is that with few exceptions, preclinical studies are conducted using healthy tissue systems. The negativism regarding purinergic therapeutics has been discussed previously [7] in relation to the universal use of aspirin and other cyclooxygenase inhibitors such as analgesics and anti-inflammatory agents. The wisdom being applied to what is known about the action of adenosine agonists in animal models would similarly preclude using aspirin because of its wide spectrum of potential actions. Yet it is fairly obvious that because of the inflammatory process, the drug acts as a paracrine effector agent to seek out diseased tissues, leaving those in normal homeostatic balance unaffected.

Another factor relates to the fact that all currently known adenosine agonists represent modified forms of the parent. This fact tends to exacerbate the potential for actions that overlap with the endogenous effector. In other successful drug discovery programs, the breakthroughs in minimizing the therapeutic ratio for deleterious side-effects has been the identification of novel structure which in turn has usually led to the identification of receptor subtypes.

### Adenosine antagonists

A similar situation exists with the xanthine adenosine antagonists. Caffeine and theophylline as the prototypic adenosine antagonists are widely ingested in foodstuffs and are probably the most avidly consumed drugs in the world. The actions of the xanthines are compounded by their activity as phosphodiesterase inhibitors. The basis for the use of theophylline in asthma has yet to be satisfactorily elucidated at the mechanistic level. Many potent and selective adenosine antagonists that have been developed (e.g. 8-substituted xanthine derivatives [1]) have yet to be introduced as therapeutic agents. Evaluation of such entities as central stimulants was disappointing [31], a fact that may be attributed to their poor bioavailability, due to hydrophobicity and highly stable crystal lattices, leading to very low (in the range of  $1 \mu M$ ) aqueous solubility. A later generation of adenosine antagonists has now been developed including the more hydrophilic xanthine amine congener (XAC), which contains an ionizable group in the form of an amine, attached through a functionalized chain at a distance from the main pharmacophore [1].

<sup>\*</sup> Macallum GE, Walker RM, Barsoum NJ and Smith GS, unpublished observations. Cited with permission.

<sup>†</sup> Trivedi BK, Bristol JA, Blankley CJ, Hamilton JW, Patt WC, Kramer WJ, Bruns RF, Cohen DM and Ryan MJ, Synthesis of prodrugs of CI-936, an adenosine receptor agonist for CNS disorders. Abstract presented to the Division of Medicinal Chemistry at the meeting of the Third Chemical Congress of North America in Toronto, 1988, Abstr. 32.

Fig. 1. Structures of two non-xanthine adenosine antagonists with selectivity for A<sub>2</sub> receptors.

In order for an adenosine antagonist to become a drug, it will have to be far superior to caffeine and theophylline, by being more selective and significantly less toxic. For this reason, it would be desirable to develop a selective, potent, and bioavailable non-xanthine adenosine antagonist. A number of classes of non-xanthine adenosine antagonists have been discovered [32]. Of these, CGS 15943A, 14 (Fig. 1) [33], was targeted in aerosol form as an antiasthmatic but failed in development due to skin irritation. A series of triazoloquinoxalines from Pfizer, which for a time were in clinical trials as antidepressants, were later found to include some A<sub>2</sub>-receptor selective adenosine antagonists [34], such as CP 66,713 (15).

One promising potential site for therapeutic application of adenosine antagonists is in the kidneys. As shown in Table 3, adenosine affects nearly all aspects of renal function: renal blood flow and its distribution within the kidney [35], glomerular filtration rate [35], renin secretion [35], urine flow and sodium excretion [36], and transmitter release from the renal efferent sympathetic nerves [37]. All of these effects are mediated by adenosine receptors, since they are antagonized by alkylxanthines and/or mimicked by adenosine analogs that act as adenosine receptor agonists [37, 38]. For some of these effects, the subclasses of adenosine receptors that are involved have been established, based on the order of potency of agonists. These observations, taken together with the observation that kidneys produce and release adenosine into extracellular fluids [39], suggest that variations in the concentration of endogenously-released adenosine could play important roles in renal function and/or dysfunction. Indeed, it has been postulated that adenosine is the mediator of several physiological and pathophysiological phenomena: the autoregulation of renal blood flow and glomerular filtration rate [35], the tubuloglomerular feedback response [40], the effect of macula densa cells on nearby reninsecreting juxtaglomerular cells [35, 36, 40], and the hemodynamic changes in acute renal failure [14]. If true, then theoretically, adenosine receptor antagonists should have equally profound and wideranging effects on renal function.

Although the concentration of endogenouslyreleased adenosine in renal extracellular fluids is in a range that suggests effects on renal hemodynamics [35], it is questionable whether variations in the concentration play a role in the normal physiological control of renal hemodynamics. There is evidence that adenosine mediates the tubuloglomerular feedback response [39, 40], but it is very unlikely to mediate autoregulation of renal blood flow and glomerular filtration rate [14]. In fact, although the renal hemodynamic effects of theophylline are extremely variable, theophylline actually improves autoregulation in some pathological circumstances [41], and in congestive heart failure, theophylline nearly doubles glomerular filtration rate in humans [42], perhaps because of unusually high levels of endogenously released adenosine. Moreover, as outlined below, there is evidence that endogenously released adenosine explains, at least in part, the renal hemodynamic changes in some experimental animal models of acute renal failure.

It was proposed [14] that adenosine mediates the hemodynamic changes in acute renal failure, and that the changes are pathogenic in reducing glomerular filtration rate. According to this hypothesis, a state of energy deficit (impaired oxidative phosphorylation, resulting from hypoxic, ischemic, or nephrotoxic tubular cell injury) leads to decreased cellular ATP levels and increased adenosine production and release. The adenosine then acts on afferent and efferent arterioles to produce the hemodynamic changes. Several observations are consistent: (a) Renal ischemia results in decreased cellular ATP levels and increased adenosine production and release. Nephrotoxic injury of renal tubular cells also decreases cellular ATP levels, but it is not known if this accompanied by increased adenosine production and release. (b) Adenosineinduced changes in renal hemodynamics (see above) mimic the hemodynamic changes in acute renal failure [14]. These include variable changes in renal blood flow and its distribution within the kidney, but consistent decreases in filtration fraction and glomerular filtration rate. (c) Pharmacological manipulation of the renal adenosine system has provided support for the hypothesis, at least in some experimental animal models, of acute renal failure. Theophylline has protective effects on renal function in an ischemic model that is produced in rats by

Table 3. Summary of the receptor-mediated\* renal effects of adenosine

	racic 5. Summary of	radic 3. Summary of the receptor-incurated remained of ademosities	
Target cells	Receptor type	Response	Comments
Vascular smooth muscle cells Afferent arteriole	A <sub>2</sub>	Constriction (decrease RBF† and GFR) Dilation (increase RBF and GFR)	RBF can decrease, increase, or remain constant but GFR always
Efferent arteriole	<b>A</b> 2	Dilation (increase RBF, decrease GFR)	uccreases. Inerc may be a redistribution of renal blood flow from outer to inner cortex. If so, urine flow and sodium excretion would decrease, since inner cortical nephrons reabsorb sodium and water very avidly.
Juxtaglomerular cells	$\mathbf{A}_2$	Inhibit renin secretion Stimulate renin secretion	Renin secretion can decrease, increase, or remain constant.
Tubular epithelial cells	Ā Ā	Decrease cyclic AMP Increase cyclic AMP	It is not clear which, if either, of these changes in cyclic AMP explain adenosine-induced decreases in urine flow and sodium excretion.
Sympathetic nerve terminals	¢.	Inhibit norepinephrine release	RBF would increase, renin secretion would decrease, and urine flow and sodium excretion would increase, since the nerves constrict (α-adrenergic), stimulate renin secretion (β-adrenergic), and enhance the reabsorption of sodium and water.

\* Adenosine-induced increases in renal afferent (sensory) nerve activity are not receptor-mediated, since they cannot be blocked by theophylline. The receptor-mediated effects listed in the table can be blocked by theophylline and are induced by adenosine analogs as well as adenosine per se.

† RBF = renal blood flow; and GFR = glomerular filtration rate.

unilateral occlusion of a renal artery for 30 or 45 min; renal plasma flow and glomerular filtration rate are higher in the previously-ischemic kidneys of rats treated with theophylline than in rats treated with the vehicle [14], during both the initiation and maintenance phases of renal failure. Moreover, the adenosine uptake blocker dipyridamole enhances the severity of failure in the initiation phase of this model [14]. Ischemia is considered to play a role in glycerol-induced myoglobinuric acute renal failure in rats, and in this model, also, theophylline has protective effects [14] that are dose-dependent and independent of any effects on sodium excretion or tubuloglomerular feedback. Bowmer et al. [43] have shown that 8-phenyltheophylline, a more potent adenosine receptor antagonist, has similar protective effects in the glycerol model, both with respect to renal function and renal morphology. Pentoxifylline [44] and theophylline [14, 43-45] have protective effects in other ischemic and toxin-induced models of renal failure in rats and rabbits. On the other hand, in other models of nephrotoxic acute renal failure [46], adenosine-mediated hemodynamic changes do appear to be less important.

Exogenous adenosine produces intense antidiuretic and antinatriuretic effects in many species [14]. These effects are receptor-mediated since they are competitively antagonized by theophylline and mimicked by several adenosine analogs. It seems reasonable to assume that the well-known diuretic and natriuretic effects of methylxanthines are produced by antagonism of the effects of endogenously released adenosine. A variety of mechanisms could be involved in adenosine-induced antidiuresis and antinatriuresis. Explanations based on systemic effects (changes in cardiac output, blood pressure, neural activity, or hormone secretion) seem to be excluded by the observations that isolated perfused kidneys respond predictably to both agonists and antagonists. However, the changes in urine flow and sodium excretion could be a consequence of a change in renal hemodynamics, since adenosine may induce a vasodilation of the juxtamedullary cortex, and it is believed that juxtamedullary nephrons reabsorb filtered water and sodium more avidly than outer cortical nephrons. In addition, adenosine decreases the glomerular filtration rate and, therefore, the filtered loads of water and sodium. However, adenosine-induced percentage decreases in urine flow and sodium excretion exceed, by far, adenosineinduced percentage decreases in glomerular filtration rate [14]. Conversely, methylxanthines can produce diuresis and natriuresis in the absence of detectable increases in glomerular filtration rate [14]. Therefore, it seems reasonable to assume that adenosineinduced antidiuresis and antinatriuresis and, by inference, methylxanthine-induced diuresis and natriuresis, can be mediated by both renal hemodynamic and direct tubular mechanisms. Consistently, adenosine analogs stimulate active sodium transport in toad kidney cells [47]. Moreover, binding studies and studies of adenylate cyclase activity demonstrate the presence of both A<sub>1</sub>- and A<sub>2</sub>-adenosine receptors [48].

As with classical adenosine agonists, a multiplicity of side-effects of potent adenosine antagonists is possible. For example, 8-phenyltheophylline causes a diabetes-like condition in the rat heart, i.e. it induces total insulin refractoriness to glucose transport [49].

## Non-classical approaches

The failure of existing entities and the perceived disadvantages of classical medicinal chemical approaches for adenosine receptor drugs has led a number of laboratories to explore alternate approaches, including prodrugs [50, \*] and indirect adenosine agonists, i.e. uptake blockers [51]. Other laboratories have turned to applications such as the treatment of stroke [8, 12], in which the potential benefit of life-saving treatment would outweigh the side-effects.

Prodrugs. A prodrug is a chemically masked derivative of an active drug, which is converted to an active form at a physiological target site [52]. The prodrug is a latent (and therefore biologically inactive or weakly active) form of a drug. The unmasking of the prodrug typically takes place through an enzymatic or a chemical cleavage step. Ideally, the free drug is present at biologically active concentrations only at the target organ or tissue. In this manner, side-effects at other sites are minimized.

A limited success was achieved when an attempt was made to separate the cardiovascular effects of CI-936 from the CNS effect by preparing a variety of prodrugs of the adenosine derivative.\* Modification of the ribose hydroxyl groups in the form of more hydrophobic esters (simple acyl and various amino acid derivatives) produced compounds which were more CNS- versus cardiovascular-selective than CI-936.

To exploit the beneficial therapeutic properties of adenosine antagonists in the kidneys, a xanthine prodrug (Fig. 2) has been developed [50]. This prodrug was based on the conceptual fusion of two developments, one being the xanthine functionalized congeners, of which XAC is the prototype, and the other a kidney prodrug scheme first proposed by Orlowski, Wilk, and coworkers [53]. The prodrug scheme utilizes selective cleavage of  $\gamma$ -glutamyl amide derivatives by the enzyme  $\gamma$ -glutamyl transpeptidase ( $\gamma$ -GT), which is highly concentrated in kidney cells of the brush border membrane. This general prodrug scheme has been expanded to achieve a selective concentration of various drugs, including dopamine and the antibiotic famethoxazole [53], in the kidney. This prodrug scheme utilizes a two-step cleavage process, in which an N-acyl-γ-glutamyl amide derivative of an aminecontaining drug is cleaved first by an acylase to give the  $\gamma$ -glutamide and finally by  $\gamma$ -GT. Both enzymes are concentrated in the kidneys. Using two enzymatic cleavage steps to generate the potent drug resulted in greater specificity for the kidney than a single step dependent on  $\gamma$ -GT. Later work has extended use of γ-glutamyl amides of diuretics as kidney prodrugs [54].

<sup>\*</sup> Hamilton HW, Hawkins LD, Patt WC, Johnson SA, Trivedi BK, Heffner TG, Wiley JN and Bruns RF, Abstract presented to the Division of Medicinal Chemistry at the 196th National ACS Meeting, 1988, Abstr. 80.

$$R = CH_{3}CO-NH CO_{2}H H (CH_{2})_{2}CO H (CH_{2})_{2}CO H (CH_{2})_{2}CO H (CH_{2})_{2}CO H H (CH_{2})_{2}CO H (CH_{2})_{2$$

Fig. 2. Enzymatic transformations of xanthine prodrugs derived from XAC, 18.

XAC, like theophylline, is active as a diuretic and as a natriuretic [50] antagonizing the reduction of glomerular filtration rate induced by the adenosine analog, N<sup>6</sup>-cyclohexyladenosine (CHA). At a dose of 5 mg/kg, administered in the tail vein in rats, XAC causes an increase in urine output that is comparable to that produced by theophylline. Furthermore, XAC is A<sub>1</sub>-selective (species dependent), and further manipulation of the receptor subtype selectivity is possible through the introduction of additional substituents apart from the amino group [1]. Thus, it would be desirable to achieve a selective concentration in the kidneys of a theophylline analog such as XAC, which is more potent and selective than theophylline. γ-Glutamyl-XAC, 17 (Fig. 2), and its derivatives were explored as prodrugs for cleavage in the kidneys by  $\gamma$ -glutamyltransferase. The basis for the expectation that y-Glu-XAC would serve as a suitably masked prodrug (and therefore less active than the parent) is the observation that, in general, anionic xanthine derivatives, e.g. functionalized on the 8-position chain, and zwitterionic derivatives tend to be less potent as adenosine antagonists than cationic (principally amine) derivatives, such as XAC and D-Lys-XAC. This generalization was substantiated in the case of glutamyl derivatives of XAC. In binding assays [50], which for adenosine antagonists are often indicative of biological potency except in cases of insolubility,

the anionic N-acetyl- $\gamma$ -glutamyl-XAC (16) was much less potent than XAC. The zwitterionic  $\gamma$ -glutamyl-XAC was intermediate in potency as an adenosine antagonist (20-fold less potent than XAC in binding assays at rat A<sub>1</sub>-adenosine receptors). It was hoped that N-acetyl- $\gamma$ -glutamyl-XAC would serve as a substrate for the two-step cleavage process. However, it was not certain that  $\gamma$ -glutamyl-XAC would be cleaved by  $\gamma$ -GT [55], although it seemed likely that N-acetyl-γ-glutamyl-XAC would be cleaved in the first step by renal acylase, known to have broad specificity. Surprisingly, N-acetyl-γ-glutamyl-XAC was not a substrate for renal acylase, and did not act as a diuretic in vivo. When N-acetyl-γ-glutamyl-XAC was injected in rats intravenously, no XAC was observed in urine or in plasma. y-Glutamyl-XAC did act as a diuretic, and was cleaved to XAC both in vivo and in pure enzyme preparations. Since the two enzyme processes [53] could not be utilized, an additional derivative, which consisted of  $\gamma$ glutamyl- $\gamma$ -glutamyl-XAC (19), which would be acted upon by  $\gamma$ -GT in two steps, was also prepared, and found to be cleavable readily to XAC. y-Glutamyl-\gamma-glutamyl-XAC was also active in vivo as a diuretic. To establish kidney selectivity of  $\gamma$ glutamyl-XAC, it will be necessary to characterize the in vivo actions of this adenosine antagonist more fully, i.e. assessing for actions in the cardiovascular system. Since even the prodrug itself potentially

Fig. 3. Structures of R 75231 (20), an analog of the adenosine uptake inhibitor, mioflazine; the binding enhancer PD 81,723 (21); and 5-amino-4-imidazole carboxamide (AICA) riboside (22).

has adenosine antagonist properties (albeit much weaker), it is possible that other biological effects will be observed. Another question which should be answered is the mechanism of entry into the cell in order to be acted upon by  $\gamma$ -GT, which occurs intracellularly.

Uptake blockers. The classical nucleoside transport inhibitor, dipyridamole, has been used clinically as a vasodilatory agent and to characterize the properties of adenosine uptake in a variety of mammalian tissues [3, 56]. Selective inhibition of adenosine uptake would act in a manner similar to the administration of adenosine agonists although conceptually the effects of such agents would be localized to tissues where adenosine-related metabolic and paracrine activities were highest. Mioflazine, a nucleoside transport inhibitor from Janssen Pharmaceutica, has proven to be efficacious as a hypnotic in animals [51] and may have anticonvulsant activity. The structure-activity relationships of various transport inhibitors have been determined, and a newer analog R 75231 [57] (20, Fig. 3) which does not cross the blood brain barrier, shows potential for cardioprotection.

Allosteric enhancers. An interesting strategy for potentially treating ischemic diseases and others that involve a "hyperproduction" of purine nucleosides is that of "allosteric" enhancers of adenosine binding [58]. As a result of a broad screening program at Warner-Lambert, a series of benzoylthiophene derivatives, including PD 81,723 (21, Fig. 3), were

discovered to cause an increase in the binding of adenosine analogs selectively at A<sub>1</sub>-adenosine receptors. Moreover, the biological effects of adenosine agonists as inhibitors of cAMP production in FRTL-5 rat thyroid cells were potentiated. Such enhancers, predicated on the discovery of more potent pharmacophores, when used therapeutically, might be expected to have biological effects only in ischemic areas where the adenosine concentration was elevated, in a more selective fashion than adenosine agonists.

Adenosine potentiators. Another means of raising the extracellular levels of adenosine, so that adenosine receptors are activated without the use of synthetic agonists, is through metabolic manipulation via "potentiator" actions [59]. Pretreatment with 5amino-4-imidazole carboxamide (AICA) riboside (22, Fig. 3) [60] causes an enhanced increase in the local adenosine concentration under conditions of myocardial ischemia in dogs. Increased blood flow, inhibition of granulocyte adherence, and less tissue injury were observed. One mechanism proposed for the cardiac protective effects of AICA riboside is an increased production of adenosine rather than inosine from ATP catabolism although the precise mechanism for this effect has yet to be determined [61].

#### ATP receptors as drug targets

While ATP represents a major potential source for adenosine, the nucleotide, together with ADP and AMP, can alter tissue function via specific cell surface recognition sites termed  $P_2$  (ATP) receptors, which are distinct from  $P_1$  (aderosine) receptors, except in the common structural features and the metabolic relationship of the endogenous ligands [62–64].

The concept of a role for ATP and related phosphorylated adenine nucleotides as mediators of cell-to-cell communication has a long and distinguished history via the efforts of Burnstock and his many coworkers over the past two decades [62]. ATP appears to be the main candidate as effector agent for the "non-cholinergic, non adrenergic" (NANC) neurotransmission in peripheral tissues. Four major types of P<sub>2</sub> receptor have been identified [63].

The  $P_{2t}$  receptor is selectively activated by 2-methylthio-ADP and is localized on platelets. It is an ADP receptor. The  $P_{2x}$  and  $P_{2y}$  receptors [62] are ATP receptors with  $\alpha,\beta$ -methylene-ATP and 2-methylthio-ATP being selective ligands, respectively, for each receptor. The  $P_{2z}$  receptor found in mast cells [63] is sensitive to ATP<sup>4-</sup>.

Activation of  $P_{2x}$  receptors causes contraction of arteries, bladder and vas deferens, whereas  $P_{2y}$  receptor activation relaxes taenia coli, trachea and vasculature, the latter effect involving nitric oxide as effector agent. Antagonists for the various  $P_2$  receptor subclasses have proven difficult to identify but  $\beta$ ,  $\gamma$ -dichloromethylene-ATP is a relative potent ( $K_i = 21 \,\mu\text{M}$ ) competitive antagonist [65]. The list of tisues [66] in which ATP receptors have been identified is growing rapidly, as has the availability of ATP analogs by which to study these receptors [64].

## Conclusion

Commercial interest in the therapeutic potential of purinergic ligands as drug entities has continued notwithstanding serious difficulties encountered by a number of large pharmaceutical companies. Attention is now turning towards non-classical approaches to purine therapeutics such as prodrugs, allosteric binding enhancement, and modulating metabolic and uptake processes as a means of avoiding the side-effects of classical agonists and antagonists. Also gaining importance as therapeutic targets for adenosine-related drugs are acute disorders, such as stroke, in which the lack of specificity for currently available adenosine pharmacophores may be acceptable in their acute use of the context of life-threatening situations. Classical competitive pharmacophores, however, should not be overlooked given recent discoveries of novel adenosine pharmacophores from natural sources [67-69].

# REFERENCES

- Jacobson KA, Adenosine (P<sub>1</sub>) and ATP (P<sub>2</sub>) receptors.
   In: Comprehensive Medicinal Chemistry (Ed. Emmett JC), Vol. 3, pp. 601-642. Pergamon Press, New York, 1990.
- Pearson JD, Ectonucleotidases. Measurement of activities and use of inhibitors. Methods Pharmacol 6: 83-108, 1985.
- 3. Stone TW, Lloyd HGE and Newby AC, Adenosine

- release. In: Adenosine and Adenosine Receptors (Ed. Williams M), pp. 173-223. Humana Press, Clifton, NJ, 1990.
- Drury A and Szent-Györgyi A, The physiological activity of adenine compounds with especial reference to their action upon the mammalian heart. J Physiol (Lond) 68: 213-237, 1929.
- Honey RM, Ritchie WT and Thompson WAR, The action of adenosine upon the human heart. Q J Med 23: 485-490, 1930.
- Medco's 1st approval—Adenocard. Scrip No. 1464: 22, 1989.
- Williams M, Adenosine receptors as drug targets: Fulfilling the promise? In: Purines in Cellular Signalling: Targets for New Drugs (Eds. Jacobson KA, Daly JW and Manganiello V), pp. 174–183. Springer, New York, 1990.
- Olsson RA and Pearson JD, Cardiovascular purinoceptors. Physiol Rev 70: 761-845, 1990.
- Heffner TG, Wiley JN, Williams AE, Bruns RF, Cougenhour LJ and Downs DA, Comparison of the behavioral effects of adenosine agonists and dopamine antagonists in mice. *Psychopharmacology* 98: 31-38, 1989.
- Bridges AJ, Moos WH, Szotek DL, Trivedi BK, Bristol JA, Heffner TG, Bruns RF and Downs DA, N<sup>6</sup>-(2,2-Diphenylethyl)adenosine, a novel adenosine receptor agonist with antipsychotic-like activity. J Med Chem 30: 1709-1711, 1987.
- Schingnitz G, Küfner-Mühl U, Ensinger H, Lehr E and Kuhn FJ, Selective A<sub>1</sub>-antagonists for treatment of cognitive deficits. *Nucleosides Nucleotides*, in press.
- Marangos PJ, Von Lubitz D, Miller LP, Daval JL and Deckert J, The central adenosine system as a therapeutic target in stroke. In: Purines in Cellular Signalling: Targets for New Drugs (Eds. Jacobson KA, Daly JW and Manganiello V), pp. 100-107. Springer, New York, 1990.
- Belardinelli L and Pelleg A, Rationale for the use of adenosine in the diagnosis and treatment of cardiac arrhythmias. In: Purines in Cellular Signalling: Targets for New Drugs (Eds. Jacobson KA, Daly JW and Manganiello V), pp. 95-99. Springer, New York, 1990.
- Churchill PC and Bidani AK, Adenosine and renal function. In: Adenosine and Adenosine Receptors (Ed. Williams M), pp. 335-380. Humana Press, Clifton, NJ, 1990.
- Gerber JG and Payne NA, The role of adenosine on the gastric acid secretory response. In: Purines in Cellular Signalling: Targets for New Drugs (Eds. Jacobson KA, Daly JW and Manganiello V), pp. 213– 219. Springer, New York, 1990.
- Howell RE, Gaudette RF, Raynor MC, Warner AT, Museham WT, Schenden JA, Perumattam J, Hiner RN and Kinnier WJ, Mechanisms of xanthine actions in models of asthma. In: Purines in Cellular Signalling: Targets for New Drugs (Eds. Jacobson KA, Daly JW and Manganiello V), pp. 370-375. Springer, New York, 1990.
- Daly JW, Adenosine receptors: Targets for future drugs. J Med Chem 25: 197-207, 1982.
- Stein HH, Prasad RN and Tietje KR, Adenosine-5'carboxamides for controlling undesired animals. U.S. Patent 4,167,565 (1979).
- Kawazoe K, Matsumato M, Tanabe S, Fujiwara M, Yanagimoto M, Hirata M and Kakiuchi K, Coronary and cardiohemodynamic effects of 2-phenylaminoadenosine (CV1808) in anaesthetized dogs and cats. Arzneimittelforschung 30: 1083-1087, 1980.
- Bruns RF, Lu GH and Pugsley TA, Characterization of the A<sub>2</sub> adenosine receptor labeled by [<sup>3</sup>H]NECA in rat striatal membranes. Mol Pharmacol 29: 331-346, 1986.

- 21. Bridges AJ, Bruns RF, Ortwine DF, Priebe SR, Szotek DL and Trivedi BK, N<sup>6</sup>-[2-(3,5-Dimethoxyphenyl)-2-(2-methylphenyl)ethyl]adenosine and its uronamide derivatives. Novel adenosine agonists with both high affinity and high selectivity for the adenosine A<sub>2</sub> receptor. J Med Chem 31: 1282-1285, 1988.
- Trivedi BK and Bruns RF, C2,N<sup>6</sup>-Disubstituted adenosines: Synthesis and structure-activity relationships. J Med Chem 32: 1667-1673, 1989.
- 23. Trivedi BK, Bridges AJ, Patt WC, Priebe SR and Bruns RF, N<sup>6</sup>-Bicycloalkyladenosines with unusually high potency and selectivity for the adenosine A<sub>1</sub> receptor. J Med Chem 32: 8-11, 1989.
- 24. Hutchison AJ, Williams M, deJesus R, Oei HH, Ghai GR, Webb RL, Zoganas HC, Stone GA and Jarvis MF, 2-Arylalkylamino-adenosine 5'-uronamides: A new class of highly selective adenosine A<sub>2</sub> receptor agonists. J Med Chem 33: 1919-1923, 1990.
- Olsson RA, Kusachi S, Thompson RD, Ukena D, Padgett W and Daly JW, N<sup>6</sup>-Substituted N-alkyladenosine-5'-uronamides: Bifunctional ligands having recognition groups for A<sub>1</sub> and A<sub>2</sub> adenosine receptors. J Med Chem 29: 1683–1689, 1986.
- 26. Klabunde RE, Dipyridamole inhibition of adenosine metabolism. Eur J Pharmacol 93: 21-26, 1983.
- Sollevi A, Cardiovascular effects of adenosine in man: Possible clinical implications. *Prog Neurobiol* 27: 319–349, 1986.
- Fredholm BB, Jacobson KA, Jonzon B, Kirk KL, Li YO and Daly JW, Evidence that a novel 8-phenylsubstituted xanthine derivative is a cardioselective adenosine receptor antagonist in vivo. J Cardiovasc Pharmacol 9: 396-400, 1987.
- Taylor DG, Bruns RF, Bjork FA, Cohen DM, Singer RM, Olszewski BJ, Ryan MJ, Trivedi BK and Kaplan HR, Antihypertensive profile of an adenosine analog PD 117,519. FASEB J 2: 1799, 1988.
- Griffin HE and Metz AL, Cardiac arrhythmias and coronary arteriopathy induced by an adenosine agonistantihypertensive in beagle dogs. J Mol Cell Cardiol 21 (Suppl II): S.45, 1989.
- Snyder SH, Katims JJ, Annau Z, Bruns RF and Daly JW, Adenosine receptors and the actions of methylxanthines. Proc Natl Acad Sci USA 78: 3260– 3264 1981
- 32. Daly JW, Hong O, Padgett WL, Shamim MT, Jacobson KA and Ukena D, Non-xanthine heterocycles: Activity as antagonists of A<sub>1</sub>- and A<sub>2</sub>-adenosine receptors. *Biochem Pharmacol* 37: 655-664, 1988.
- 33. Francis JE, Cash WD, Psychoyos S, Ghai G, Wenk P, Friedmann RC, Atkins C, Warren V, Furness P, Hyun JL, Stone GA, Desai M and Williams M, Structure activity profile of a series of novel triazoloquinazoline adenosine agonists. J Med Chem 31: 1014–1020, 1988.
- 34. Sarges R, Howard HR, Browne RG and Koe BK, 4-Amino [1, 2, 4]triazolo[4, 3 a]quinoxalines Highly selective adenosine and potential antidepressants. In: Purines in Cellular Signalling: Targets for New Drugs (Eds. Jacobson KA, Daly JW and Manganiello V), pp. 417-418. Springer, New York, 1990.
- Spielman WS and Thompson CI, A proposed role for adenosine in the regulation of renal hemodynamics and renin release. Am J Physiol 242: F423-F435, 1982.
- Tagawa H and Vander AJ, Effects of adenosine compounds on renal function and renin secretion in dogs. Circ Res 26: 327-338, 1970.
- Hedqvist P, Fredholm BB and Olundh S, Antagonistic effects of theophylline and adenosine on adrenergic neuroeffector transmission in the rabbit kidney. Circ Res 43: 592-598, 1978.
- Murray RD and Churchill PC, The concentrationdependency of the renal vascular and renin secretory

- responses to adenosine receptor agonists. *J Pharmacol Exp Ther* 232: 189–193, 1985.
- Osswald H, Schmitz H-J and Kemper R, Tissue content of adenosine, inosine and hypoxanthine in the rat kidney after ischemia and postischemic recirculation. *Pflugers Arch* 371: 45-49, 1977.
- Osswald H, Nabakowski G and Hermes H, Adenosine as a possible mediator of metabolic control of glomerular filtration rate. *Int J Biochem* 12: 263-267, 1980.
- 41. Osborn JE, Hoversten LG and DiBona GF, Impaired blood flow autoregulation in nonfiltering kidneys: Effects of theophylline administration. *Proc Soc Exp Biol Med* 174: 328-335, 1983.
- 42. Davis JO and Shock NW, The effect of theophylline ethylene diamine on renal function in control subjects and in patients with congestive heart failure. J Clin Invest 28: 1459-1468, 1949.
- 43. Bowmer CJ, Collis MG and Yates MS, Effect of the adenosine antagonist 8-phenyltheophylline on glycerolinduced acute renal failure in the rat. *Br J Pharmacol* 88: 205–212, 1986.
- 44. Ellermann J, Grunder W and Keller T, Effect of pentoxifylline on the ischemic rat kidney monitored by <sup>31</sup>P NMR spectroscopy in vivo. Biomed Biochim Acta 47: 515-521, 1988.
- 45. Gouyon J-B and Guignard J-P, Functional renal insufficiency: Role of adenosine. *Biol Neonate* **53**: 237–242, 1988.
- Rossi N, Ellis V, Kontry T, Gunther S, Churchill P and Bidani A, The role of adenosine in HgCl<sub>2</sub>-induced acute renal failure in rats. Am J Physiol 258: F1554– F1560, 1989.
- Lang MA, Preston AS, Handler JS and Forrest JN Jr, Adenosine stimulates sodium transport in kidney A6 epithelia in culture. Am J Physiol 249: C330-C336, 1985.
- Spielman WS, Arend LJ, Klotz K-N and Schwabe U, Adenosine receptors and signaling in the kidneys. In: Purines in Cellular Signalling: Targets for New Drugs (Eds. Jacobson KA, Daly JW and Manganiello V), pp. 220-225. Springer, New York, 1990.
- Law WR, McLane MP and Raymond RM, Adenosine is required for myocardial insulin responsiveness in vivo Diabetes 37: 842-845, 1988.
- Barone S, Churchill PC and Jacobson KA, Adenosine receptor prodrugs: Towards kidney-selective dialkylxanthines. J Pharmacol Exp Ther 250: 79–85, 1989.
- Wauquier A, Van Belle H, Van der Broeck WAE and Janssen PAJ, Sleep improvement in dogs after oral administration in mioflazine, a nucleoside transport inhibitor. *Psychopharmacology* 91: 434-440, 1987.
- Bundgaard H, Design of prodrugs: Bioreversible derivatives for various functional groups and chemical entities. In: *Design of Prodrugs* (Ed. Bundgaard H), pp. 1-92. Elsevier, Amsterdam, 1985.
- Orlowski M, Mizoguchi H and Wilk S, N-Acyl-γglutamyl derivatives of sulfamethoxazole as models of kidney-selective prodrugs. J Prodrug Exp Ther 212: 167-172, 1980.
- 54. Hoffbauer KG, Sonnenburg C, Stalder R, Criscione L, Kraetz J, Fuhrer W and Habicht E, GP 22979A, a renal vasodilator with natriuretic properties. J Pharmacol Exp Ther 232: 838-844, 1985.
- Magnan SDJ, Shirota FN and Nagasawa HT, Drug latentiation by γ-glutamyl transpeptidase. J Med Chem 25: 1018–1021, 1982.
- 56. Marangos PJ, Deckert J and Bisserbe J-C, Central sites of adenosine action and their interaction with various drugs. In: Topics and Perspectives in Adenosine Research (Eds. Gerlach E and Becker BF), pp. 74-88. Springer, Berlin, 1987.

- 57. van Belle H, Comparative pharmacology of nucleoside transport inhibitors. *Nucleosides Nucleotides*, in press.
- 58. Bruns RF and Fergus JH, Allosteric enhancers of adenosine A<sub>1</sub> receptor binding and function. In: Adenosine Receptors in the Nervous System (Ed. Ribeiro JA), pp. 53-60. Taylor & Francis, London, 1989.
- Engler R, Consequences of activation and adenosinemediated inhibition of granulocytes during myocardial ischemica. Fed Proc 46: 2407, 1987.
- 60. Gruber HA, Hoffer ME, McAllister DR, Laikind PK, Lane TA, Schmid-Schoenbein GW and Engler RL, Increased adenosine concentration in blood from ischemic myocardium by AICA riboside. Effects on flow, granulocytes, and injury. Circulation 80: 1400– 1411, 1989.
- 61. Mullane KM and Williams M, Adenosine and cardiovascular function: In: Adenosine and Adenosine Receptors (Ed. Williams W), pp. 289-319. Humana Press, Clifton, NJ, 1990.
- 62. Burnstock G and Kennedy C, Is there a basis for distinguishing two types of P<sub>2</sub>-purinoceptors? *Gen Pharmacol* 16: 443, 1985.
- 63. Gordon JL, Extracellular ATP: Effects, sources, and fate. *Biochem J* 233: 309-319, 1986.

- 64. Williams M and Cusack NJ, Neuromodulatory roles of purine nucleosides and nucleotides: Their receptors and ligands. *Neurotransmissions* 6: 1-6, 1990.
- 65. Krishtal OA, Marchenko SM, Obukhov AG and Volkova TM, Receptors for rat sensory neurons: The structure-function relationships for ligands. Br J Pharmacol 95: 1057-1062, 1988.
- Burnstock G, Classification and characterization of purinoceptors. In: Purines in Cellular Signalling: Targets for New Drugs (Eds. Jacobson KA, Daly JW and Manganiello V), pp. 241–253. Springer, New York, 1990.
- 67. Davies LP, Jamieson DD, Baird-Cambert JA and Kazlauskas R, Halogenated pyrrolopyrimidine analogues of adenosine from marine organisms: Pharmacological activities and potent inhibition of adenosine kinase. *Biochem Pharmacol* 33: 347-355, 1984.
- 68. Balduini W and Cattabeni F, Displacement of [3H]N<sup>6</sup>-cyclohexyladenosine binding to rat cerebral cortical membranes by a hydroalcoholic extract of Valeriana officinalis. Med Sci Res 17: 639-640, 1989.
- 69. Araki H, Karasawa Y, Kawashima K, Hayashi M, Aihara H and Jun-Hua H, The adenosine analogue and cerebral protecting agent AMG-1 has no effect on delayed neuronal death following ischemia. *Methods Find Exp Clin Pharmacol* 11: 731-736, 1989.